



## Topsentiasterol sulfates with novel iodinated and chlorinated side chains from the marine sponge *Topsentia* sp.

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### ABSTRACT

Three new marine polar steroids, the first chlorine-containing steroid sulfate (**1**), topsentiasterol sulfate F (**2**) and the first natural iodinated steroid (**3**) have been isolated from the marine sponge *Topsentia* sp. The structures of **1–3** were elucidated using NMR and HRESIMS as well as by chemical correlation of **1** with previously known topsentiasterol sulfate D. Compound **1** proved to be an effective inhibitor of *endo*-1,3- $\beta$ -D-glucanase from the marine mollusc *Spisula sachalinensis*.

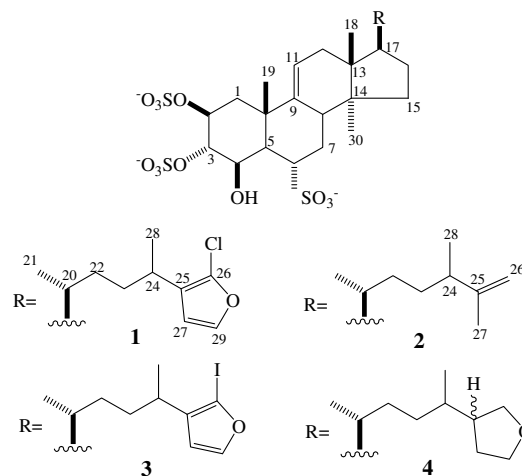
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Although halogen-containing secondary metabolites are well known and are abundant in marine organisms, only a few iodinated natural products<sup>1–5</sup> or chlorinated steroids<sup>6–11</sup> are known at the present time.

In continuation of our search for new physiologically active marine natural products,<sup>12–14</sup> we have studied topsentiasterol sulfates isolated from a sponge belonging to the genus *Topsentia* (family Halichondriidae, class Demospongiae) collected from Vietnamese waters (depth 10–15 m, Vang Fong Bay 12°35,993N 109°18,596E, June 2007). As a result, a new chlorine-containing steroid named as chlorotopsentiasterol sulfate D (**1**)<sup>15</sup> along with the earlier unknown topsentiasterol sulfate F (**2**)<sup>16</sup> and a unique iodinated steroid (**3**)<sup>17</sup> were isolated and structurally elucidated. Herein we report the structures of **1–3**.

The ethanol extract of the frozen sponge (dry weight 67 g) was concentrated and subjected to column chromatography on Polychrome-1 (powdered Teflon) using H<sub>2</sub>O, followed by silica gel chromatography (CHCl<sub>3</sub>–EtOH–H<sub>2</sub>O, 100:125:25) to obtain a crude mixture of topsentiasterol sulfates. This was further separated by preparative HPLC (YMC-ODS-A column, 80:20:1% MeOH–H<sub>2</sub>O–1 M CH<sub>3</sub>COONH<sub>4</sub>) to give a fraction of halogenated topsentiasterol sulfates (38.8 mg) and **2** (0.3 mg, 0.0004% of dry weight) along with previously known topsentiasterol sulfates D<sup>18</sup> (23.3 mg, 0.03% of

dry weight) and E<sup>18</sup> (3.5 mg, 0.005% of dry weight). The fraction containing halogenated topsentiasterol sulfates was twice rechromatographed by HPLC (YMC-ODS-A column, 75:25:1% MeOH–H<sub>2</sub>O–1 M CH<sub>3</sub>COONH<sub>4</sub>) affording 0.5 mg of the subfraction, containing iodotopsentiasterol sulfate D (**3**), contaminated with chlorotopsentiasterol D (**1**) and pure **1** itself (15.6 mg, 0.02% of dry weight).



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NMR data of the isolated polar steroids (Table 1) showed that these compounds belong to the 4 $\beta$ -hydroxy-14 $\alpha$ -methyl steroid trisulfates, discovered by Fusetani's group from *Topsentia* sp. collected at Ishigaki Island, Okinawa.<sup>18</sup> Topsentiasterol sulfates A–E,<sup>18</sup> Sch 55867,<sup>19</sup> and spheciosterol sulfates A–C<sup>20</sup> are the only previously reported members of the 4 $\beta$ -hydroxy-14 $\alpha$ -methyl steroid group isolated from sponges.<sup>20</sup> Similar to the related ibisterol,<sup>21</sup> these unusual steroid polysulfates contain a  $\Delta^{9(11)}$ -unsaturated steroid nucleus and sulfate groups at the 2 $\beta$ , 3 $\alpha$ , and 6 $\alpha$  positions. <sup>1</sup>H and <sup>13</sup>C NMR spectra of **1** were similar to those of other topsentiasterol sulfates,<sup>18</sup> including the signals of three angular methyl groups, a C-9(11)-double bond, and four oxygenated methines. These NMR spectra differed from those of topsentiasterol sulfate D mainly in the absence of a broad singlet, H-26, of a 3-substituted furan moiety. Consequently, the structure **1** was suggested as an analogue of topsentiasterol sulfate D with modification at C-26, and subsequently confirmed by analysis of COSY, DEPT, HSQC, and HMBC data (Fig. 1).

**Table 1**

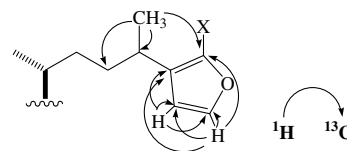
NMR data for chlorotopsentiasterol sulfate D (**1**) and topsentiasterol sulfate F (**2**) in CD<sub>3</sub>OD

Position	<b>1</b>		<b>2</b>		COSY
	$\delta_{\text{H}}$ (mult, J in Hz) <sup>a</sup>	COSY	$\delta_{\text{C}}^{\text{b}}$	$\delta_{\text{H}}$ (mult, J in Hz)	
1a	1.82 dd (3.7; 14.6)	H1b, H2	38.0	1.86 m	H1b, H2
1b	2.40 br d (14.6)	H1a		2.28 br d (13.9)	H1a
2	4.95 m	H1a, H1b, H3	76.3	4.94 m	H1a
3	4.74 dt (2.6; 1.0)	H2, H4, H1b	76.6	4.77 m	
4	4.48 dt (2.8; 1.2)	H3, H5, H6	69.2	4.45 m	H5
5	1.51 dd (2.9; 11.4)	H4, H6	48.6	1.46 m	H4, H6
6	4.81 dd (4.5; 11.3)	H5, H7a, H7b	77.1	4.81 m	H7a, H7b
7a	1.56 m	H6, H7b, H8	36.0	1.56 m	H6, H7b, H8
7b	2.21 dt (5.1; 11.7)	H6, H7a, H8		2.22 dt (4.3; 12.4)	H6, H7a, H8
8	2.48 m	H7a, H7b	42.0	2.49 m	H7a, H7b
9	—		147.0	—	
10	—		40.1	—	
11	5.34 m	H12a, H12b	118.2	5.35 m	H12a
12a	2.11 br d (17.2)	H12b	38.9	2.12 br d (17.2)	H12b
12b	1.97 m	H11, H12a		1.96 m	H11, H12a
13	—		46.2	—	
14	—		48.4 <sup>c</sup>	—	
15a	1.36	H15b	35.4	1.28 m	
15b	1.42 m	H15a		1.28 m	
16a	1.25 m	H16b, H17	29.4	1.40 m	
16b	1.85 m	H15a, H15b, H16a		1.90 m	
17	1.63 m	H16a, H16b	52.8	1.65 m	
18	0.67 s		15.6	0.70 s	
19	1.43 s		26.0	1.42 s	
20	1.36 m	H17, H21	37.9	1.36 m	H21
21	0.89 d (6.5)	H20	19.6	0.90 m	H20
22a	1.03 m	H22b	35.8	0.87 m	
22b	1.37 m	H21, H22a		1.32 m	
23a	1.36 m	H22a, H23b	35.5	1.14 m	
23b	1.60 m	H23a		1.45 m	
24	2.56 m	H23a, H28	32.2	2.05 m	H28
25	—		133.5	—	
26	—	H27	126.4	4.66 s	H27
27	6.39 d (2.0)	H26	112.4	1.65 br s	H26
28	1.14 d (7.0)	H24	21.7	1.00 d (6.9)	H24
29	7.36 d (2.0)		143.5	—	
30	0.80 s	H18	19.4	0.81 s	

<sup>a</sup> Recorded at 500 MHz.

<sup>b</sup> Recorded at 125 MHz.

<sup>c</sup> Assignment made by HMBC.



X=H Topsentiasterol sulfate D; X=Cl Chlorotopsentiasterol sulfate D

**Figure 1.** Partial structure of the side chain in topsentiasterol sulfate D and **1** with selected HMBC correlations.

The molecular formula C<sub>30</sub>H<sub>45</sub>O<sub>14</sub>S<sub>3</sub>Cl of **1** was obtained from HRESIMS measurement of the ion peaks [M<sub>3H</sub>H] (m/z 759.1575;  $\Delta$  0.8 ppm) and [M<sub>3H</sub>-H-SO<sub>3</sub>]<sup>2-</sup> (m/z 339.0999;  $\Delta$  5.1 ppm). This steroid showed MS isotopic patterns characteristic of monochlorinated compounds ([M<sub>3H</sub>-H]<sup>-</sup>:([M<sub>3H</sub>-H]+2)<sup>-</sup> = 3:1). All other structural features of **1** were supported by COSY, DEPT, HSQC, and HMBC data. To confirm that **1** is a derivative of topsentiasterol sulfate D, the chemical correlation of **1** with topsentiasterol sulfate D was carried out. Both **1** and topsentiasterol sulfate D were hydrogenated over Adams' catalyst to give the same tetrahydrofuran derivative **4** identified by NMR and HRESIMS data.<sup>22</sup>

It is of special interest that chlorotopsentiasterol sulfate D (**1**) contains a chlorofuran fragment. This structural feature had never been found previously in natural products.<sup>23</sup>

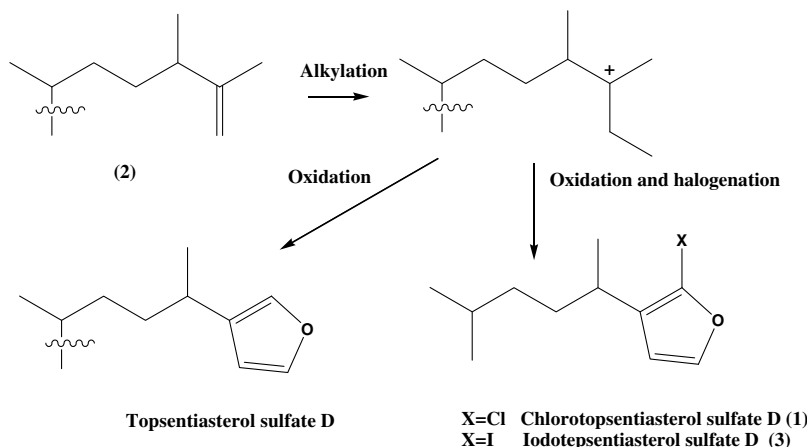
Another new steroid, named topsentiasterol sulfate F (**2**) had the molecular formula of C<sub>29</sub>H<sub>48</sub>O<sub>13</sub>S<sub>3</sub> as determined by HRESIMS measurement of the ion peak [M<sub>3H</sub>-H] (m/z 699.2169;  $\Delta$  1.4 ppm). Interpretation of the <sup>1</sup>H NMR and COSY spectra indicated that the steroid nucleus of **2** is identical to that of topsentiasterol sulfates. The remaining signals corresponded well with those reported for the 24-methyl-25(26)-ene side chains of codisterol from the marine alga *Codium fragila*<sup>24</sup> and halistanol sulfate H from the marine sponge *Pseudoaxinissa digitata*.<sup>25</sup> On the basis of all the above-discussed data, the structure of topsentiasterol sulfate F was established as **2**.

We have partly separated compound **3** from the fraction contaminated with **1**. This compound was identified as an analogue of topsentiasterol sulfate D with iodination at C-26 by <sup>1</sup>H NMR and MS. Compound **3** had the molecular formula C<sub>30</sub>H<sub>45</sub>O<sub>14</sub>S<sub>3</sub>I as determined by HRESIMS measurement of the ion peaks [M<sub>3H</sub>-H]<sup>-</sup> (m/z 851.0970;  $\Delta$  3.0 ppm.) and [M<sub>3H</sub>-H-SO<sub>3</sub>]<sup>2-</sup> (m/z 385.0668;  $\Delta$  3.9 ppm). Its NMR spectra were closely related to those of **1** differing mainly in chemical shifts in the furan moiety. To the best of our knowledge, iodotopsentiasterol sulfate (**3**) is the first example of a natural iodine-containing steroid.

It may be that topsentiasterol sulfates with unusual side chains such as in **1**, **3**, and topsentiasterol sulfate D are biosynthesized from a sterol (or sterol polysulfate) precursor having side chains of the codisterol type, for example **2**. This biosynthesis would be realized through an additional alkylation followed by oxidation and, in the case of **1** and **3**, halogenations (Scheme 1).

Steroids related to topsentiasterol sulfates demonstrate diverse biological activities, including anti-HIV, antibacterial and antifungal properties. For example, early findings on the biological properties of topsentiasterol sulfates included antibacterial activity against *Pseudomonas aeruginosa* and *Escherichia coli*,<sup>18</sup> and antifungal activity (steroid Sch 575867 from a marine sponge of the family Astroscleridae).<sup>19</sup> Spheciosterol sulfates A–C from *Spheciostorgia* sp. inhibited PKC $\zeta$ .<sup>20</sup>

We have studied the effects of **1**, **2**, and topsentiasterol sulfates D and E as possible inhibitors of 1,3- $\beta$ -glucanases.<sup>26</sup> All the substances studied did not influence the activity of exo-1,3- $\beta$ -D-glucanase from the marine filamentous fungus *Chaetomium indicum*,<sup>29</sup> but appeared to be inhibitors of endo-1,3- $\beta$ -D-glucanase from the marine mollusc *Spisula sachalinensis*.<sup>30</sup> The efficiency of endo-1,3- $\beta$ -D-glucanase inhibition was found to depend on the



**Scheme 1.** Hypothetical pathways for the biosynthesis of the side chains in furan-containing topsentiasterol sulfates.

structural peculiarities of the compounds tested. Compounds **1**, **2**, and topsentiasterol sulfates D and E inhibited *endo*-1,3- $\beta$ -D-glucanase from *S. sachalinensis* with IC<sub>50</sub> values of 5.4, 49.6, 12.4, and 7.4  $\mu$ M/L, respectively.

Thus, topsentiasterol sulfates represent a new structural group of *endo*-1,3- $\beta$ -D-glucanase inhibitors. Earlier, other polyhydroxysteroidal inhibitors of these enzymes were found.<sup>31,32</sup> Future studies will show whether the molecular actions of these two inhibitory groups are similar.

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### Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.tetlet.2008.10.007.

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- Chlorotopsentiasterol sulfate D (1): white solid; [ $\alpha$ ]<sub>D</sub><sup>20</sup> +46 (c 0.09, MeOH); <sup>1</sup>H, <sup>13</sup>C NMR data, Table 1; HRESIMS *m/z* 759.1575 [M<sub>3H</sub>-H]<sup>-</sup> (calcd for C<sub>30</sub>H<sub>44</sub>O<sub>4</sub>S<sub>3</sub>Cl 759.1582).
- Topsentiasterol sulfate F (2): white solid; {[ $\alpha$ ]<sub>D</sub><sup>20</sup> +40 (c 0.03, MeOH); <sup>1</sup>H NMR data, Table 1; HRESIMS *m/z* 699.2169 [M<sub>3H</sub>H]<sup>-</sup> (calcd for C<sub>29</sub>H<sub>47</sub>O<sub>13</sub>S<sub>3</sub> 699.21788).
- Iodotopsentiasterol sulfate D (3): selected <sup>1</sup>H NMR data (500 MHz, CD<sub>3</sub>OD): 0.68 (3H, s, H-18); 0.81 (3H, s, H-30), 0.89 (3H, d, *J* = 6.4, H-21), 1.13 (3H, d, *J* = 7.0, H-28), 1.43 (3H, s, H-19), 1.85 (1H, m, H-1a), 2.22 (1H, m, H-7b), 2.32 (1H, br d, *J* = 14.4, H-1b), 2.48 (1H, m, H-8), 5.34 (1H, m, H-11), 6.32 (0.8H, d, *J* = 2.0, H-27, for 3), 6.39 (0.2H, d, *J* = 2.0, H-27, for 1), 7.60 (0.8H, d, *J* = 2.0, H-29 for 3), 7.36 (0.2H, d, *J* = 2.0, H-29 for 1); HRESIMS *m/z* 851.0970 [M<sub>3H</sub>-H]<sup>-</sup> (calcd for C<sub>30</sub>H<sub>44</sub>O<sub>14</sub>S<sub>3</sub>I 851.0943).
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- Compound 4.** PtO<sub>2</sub> was added to a solution of **1** (2.0 mg) or topsentiasterol sulfate D (3.0 mg) in MeOH (2 ml) and stirred under H<sub>2</sub> at 25 °C for 12 h. Removal of the catalyst by filtration and evaporation of the solvent gave **4** (2.0 mg) from **1** and 3.0 mg from topsentiasterol sulfate D. Selected <sup>13</sup>C NMR (125 MHz, CD<sub>3</sub>OD) 15.6 (C18), 18.9/18.7 (C28), 19.4 (C30), 26.0 (C19), 35.4 (C22), 36.0 (C7), 37.9 (C20), 38.1 (C1), 38.9(C12), 40.0 (C10), 42.0 (C8), 46.2 (C13), 47.7/47.6 (C25), 48.6 (C5), 52.9 (C17), 69.2 (C4), 70.0/69.9 (C29), 73.8/73.6 (C26), 76.5 (C2), 76.6 (C3), 77.2 (C6), 118.3 (C11), 147.0 (C9); HRESIMS: *m/z* 729.2292 [M<sub>3H</sub>-H]<sup>-</sup> (calcd for C<sub>30</sub>H<sub>49</sub>O<sub>14</sub>S<sub>3</sub> 729.2295).
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- Determination of inhibiting activity.** The standard reaction mixture, containing 20  $\mu$ l of enzyme (1  $\times$  10<sup>-2</sup> units) and 20  $\mu$ l of inhibitor solutions (1–20  $\mu$ g of substance in water) was incubated at 25 °C for 10 min. After adding 480  $\mu$ l of a substrate<sup>27</sup> (Laminaran, 1 mg/ml), the mixture was incubated again at 37 °C for 15 min. Residual activity of the enzyme was estimated by the measurement of reducing sugars using the method of Nelson.<sup>28</sup>
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